

POLYMER SUPPORT OLIGONUCLEOTIDE SYNTHESIS VI¹⁻⁵⁾
 USE OF INORGANIC CARRIERS

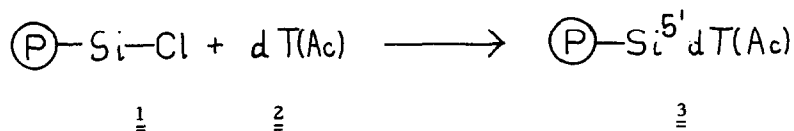
H. Köster

Institut für Organische Chemie und Biochemie, Universität Hamburg
 2 Hamburg 13, Papendamm 6, Germany

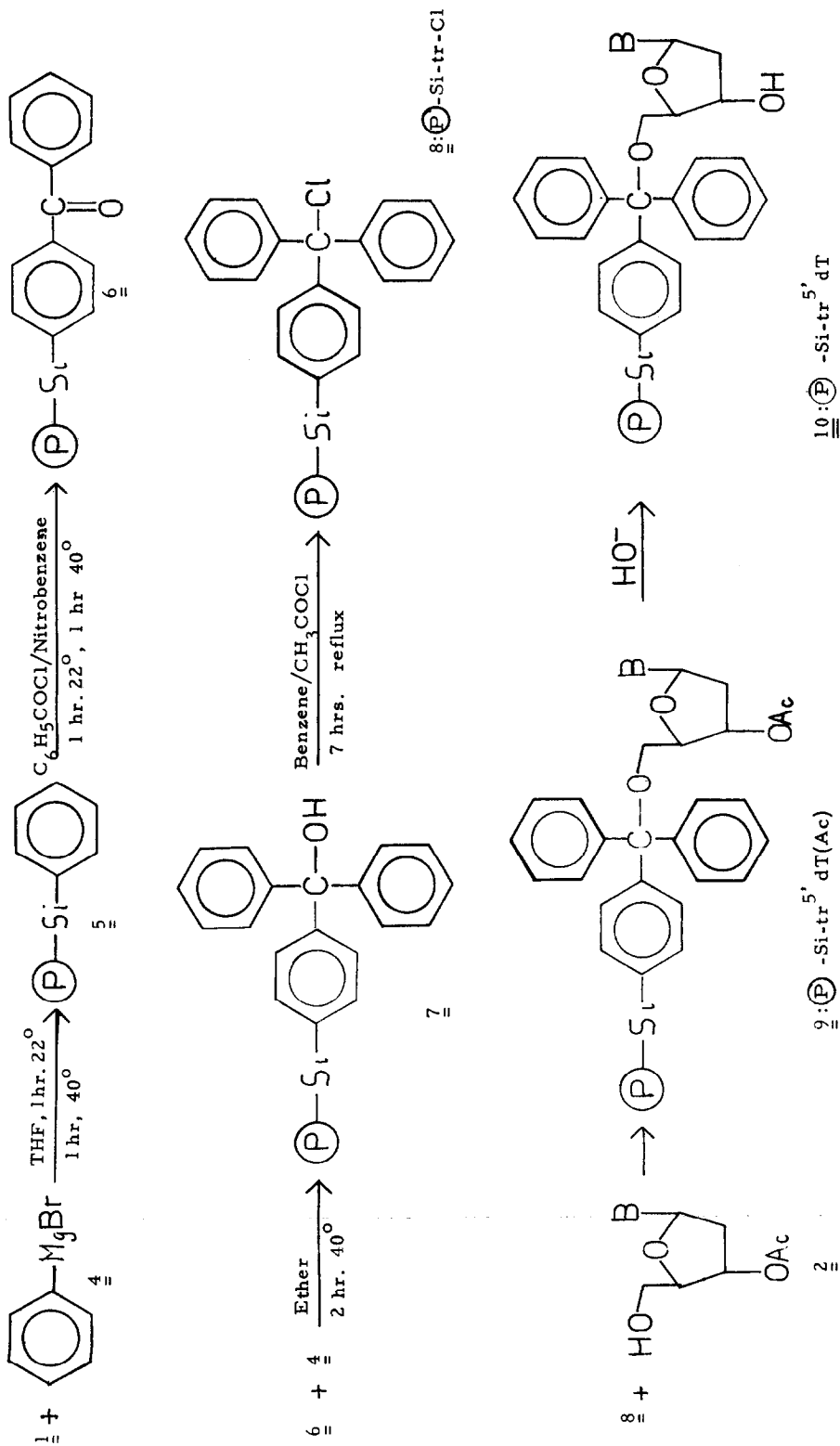
(Received in UK 28 February 1972; accepted for publication 8 March 1972)

Various methods for the synthesis of oligonucleotides on polymeric carriers are in progress in this laboratory. We first used makroporous, nonswellable polystyrene^{1, 3, 5)} and were able to synthesize dT(pdT)₇ and dTpdTpdApdCpdCpdTpdA^{4, 6)}. In another study we used popcorn polystyrene for the synthesis of dT(pdT)₅²⁾. In testing hydrophobic polymeric carriers, which are swellable in organic solvents such as popcorn polystyrene²⁾, we observed some difficulties when synthesizing an oligonucleotide chain due to the highly polar phosphodiester linkage. We therefore decided to look for hydrophylic polymeric carriers⁷⁾ in order to overcome this problem.

In a first attempt we tried nonporous glass beads (0.02 - 0.05 mm in diameter; surface $\ll 1 \text{ m}^2/\text{g}$) which would allow the oligonucleotide chain to be attached only on to the outer surface of the spheres. The glass beads could be condensed with pU(Ac)₂ using triisopropylbenzene sulfonylchloride (TIPS) in pyridine (4 hrs. 22°) or could be converted with thionyl chloride/benzene (1 : 2, v/v, reflux for 4 hrs.)⁸⁾ into the Si-Cl-derivative 1. This could be loaded with dT(Ac) (2) in pyridine (20 hrs., 22°). In both cases only 0.1 μmol pU(Ac)₂ or dT(Ac) (2) could be bound per g carrier due to the very small sur-



face area. Therefore we used silica gel (Merck, $< 0.08 \text{ mm}$ in diameter, $427 \text{ m}^2/\text{g}$ surface area). This could be converted to the Si-Cl-derivative 1 ($116 \mu\text{mol Cl}^-/\text{g}$), which reacted with dT(Ac) (2) (pyridine, 12 hrs., 22°) in a yield of $5.5 \mu\text{mol}/\text{g}$. Direct condensation of silica gel and pdT(Ac) using TIPS (1 g silica gel, 0.2 mmol pdT(Ac), 0.5 mmol TIPS, 3 hrs., 22°) gave $\textcircled{\text{P}}\text{-Si-pdT}(\text{Ac})$ in a yield of $45.5 \mu\text{mol}/\text{g}$. In both cases the nucleotidic compounds could be cleaved off either by 0.1 N hydrochloric acid or 0.1 N sodium hydroxide, the reaction being complete within 20 minutes by 22°.



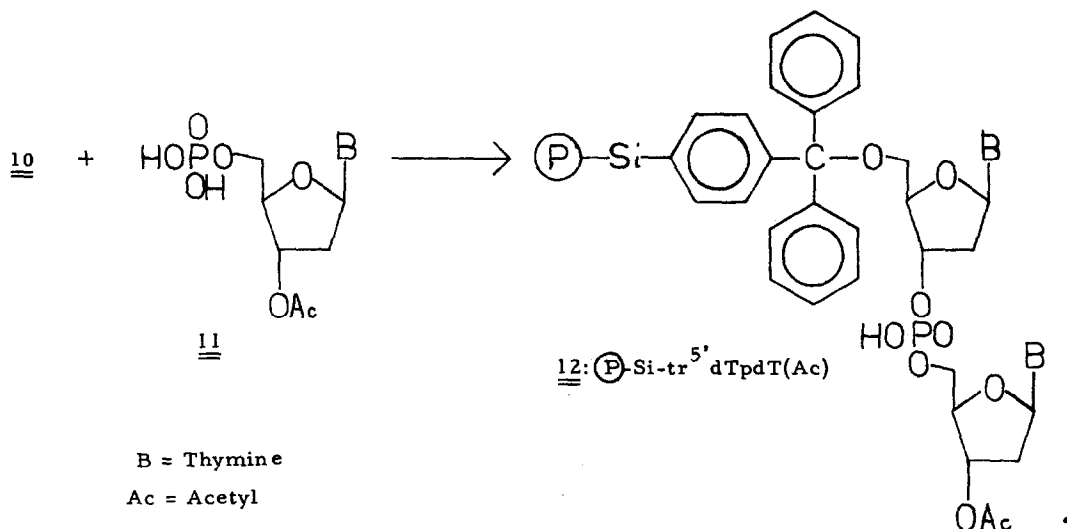
Ⓟ = polymeric backbone

B = Thymine

Ac = Acetyl

In consequence of the lability of the Si-O-P- and Si-O-C-linkage, which make them unsuitable for oligonucleotide synthesis⁹⁾, we converted 1 via the reaction sequence shown into a silica gel carrying a trityl carbinol group (7) which is bound by the more stable Si-C-linkage. By treatment with benzene/acetic acid chloride (reflux, 7 hrs.) 7 can be converted to a polymeric bound trityl chloride group (8) (0.19 mmol Cl⁻/g). In a coupling reaction with dT(Ac) (2) 10.7 μmol dT(Ac) could be bound to the carrier 8 to give (P)-Si-tr^{5'}dT(Ac) (9).

Cleavage off the carrier 9 can be accomplished using 80% acetic acid (2 hrs., 70°); no more could be cleaved off with 1 N hydrochloric acid (2 hrs., 22°). Under alkaline conditions (0.1 sodium hydroxide, 1 hr., 22°) no nucleotidic material could be cleaved off the carrier 9, however deacetylation was shown by thin layer chromatography (Merck silica gel, chloroform/methanol = 9 : 1, v/v) to be quantitative.



Condensation of 10 with pdT(Ac)(11) in pyridine (0.2 g 10, 0.1 mmol 11, 0.25 mmol TIPS, 10 hrs., 20°) gave dTpdT(Ac) in a yield of 54%, along with pdT(Ac) which had reacted with the Si-OH-groups and which was also split off during treatment with 80% acetic acid. This unwanted side reaction can be prevented by protecting the free silanol groups with p-bromomethylphenyl-trichlorosilane¹⁰⁾ before the conversion of 6 to 7. For the synthesis of longer oligonucleotides by this method including the purine nucleotides it might be advantageous to introduce the mono- or dimethoxytrityl chloride group rather than the trityl chloride group in consideration of the enhanced acid lability and to use infusorial earth in consequence of larger pores¹¹⁾ (surface area 4.2 m²/g, average pore diameter 11,000 Å, and a small distribution of pore radii) or Bio-glass 2500 (Bio-Rad Laboratories). Work along these lines are in progress.

Acknowledgement :

The author wishes to thank Mrs. C. Woldtman for her expert assistance and the Deutsche Forschungsgemeinschaft for financial support

References :

- 1) Part I in this series
F. Cramer and H. Köster , Angew. Chem. 80, 488 (1968) ;
Angew. Chem. intern. Edit. 7 , 473 (1968)
- 2) Part II in this series
H. Köster and F. Cramer , in preparation
- 3) Part III in this series
H. Köster and F. Cramer, in preparation
- 4) Part IV in this series
H. Köster, F. Cramer and A. Pollak, in preparation
- 5) Part V in this series
H. Köster and F. Cramer, in preparation
- 6) Abbreviations according to IUPAC-IUB Recommendations,
Eur. J. Biochem. 15, 203 (1970) ;
T = Thymidine, U = Uridine , d = deoxy , pdT = Deoxythymidine-5'-phosphate,
dTp = Deoxythymidine-3'-phosphate, dT(Ac) = 3'-O-Acetyl-deoxythymidine,
(P)- = Polymeric carrier
- 7) H. Köster and K. Heyns, Tetrahedron Letters 1972 , following paper
- 8) H. Deuel, J. Wartmann, K. Hutschneker, U. Schobinger and C. Güdel,
Helvetica Chimica Acta XLII, 1160 (1959)
- 9) E. Bayer, G. Jung, I. Halasz and I. Sebastian, Tetrahedron Letters 1970 , 4503
- 10) W. Parr and K. Grohmann, Tetrahedron Letters 1971 , 2633
- 11) H. L. Ritter and L. C. Drake, Ind. Eng. Chem. Anal. Edit. 17 , 787 (1945)